

Apoptosis, Fas and systemic autoimmunity: the MRL-*lpr/lpr* model

Gary G Singer, Ana C Carrera, Ann Marshak-Rothstein,
Carlos Martínez-A and Abul K Abbas

Brigham and Women's Hospital and Harvard Medical School, Boston, USA, Universidad
Autónoma de Madrid, Madrid, Spain and Boston University School of Medicine, Boston, USA

Proteins encoded by the *fas* and *fas ligand (fasL)* genes are involved in apoptotic cell death in lymphocytes. In this article we review the recent elucidation of the role of the Fas-FasL interactions in the maintenance of tolerance to self antigens and in the homeostatic regulation of lymphocyte clonal expansion, and discuss the mechanisms of autoimmunity in Fas- and FasL-deficient mutant mouse strains.

Current Opinion in Immunology 1994, 6:913-920

Introduction

The death of lymphocytes by apoptosis is an important mechanism for maintaining self-tolerance and homeostasis in the immune system. Failure of apoptotic cell death has long been postulated as a cause of loss of self-tolerance and the development of autoimmunity. Mice expressing the *lpr* (lymphoproliferation) and *gld* (generalized lymphoproliferative disease) mutations develop an accelerated, lupus-like systemic autoimmune disease. The findings that these single-gene defects are due to abnormalities in the cell death associated gene, *fas*, or in the Fas ligand (FasL), have formally established the importance of apoptotic cell death in preventing autoimmunity. These two animal models represent the first autoimmune diseases in which the major underlying genetic abnormality is known. In this paper, we review the role of apoptosis in the maintenance of self-tolerance, the genetic and immunologic basis of disease in MRL-*lpr/lpr* mice, and the relevance of this model in other animal and human models of systemic autoimmunity.

Lymphocyte apoptosis and tolerance to self antigens

Apoptotic cell death is characterized by fragmentation of DNA into nucleosome-sized fragments, nuclear dissolution and cell shrinkage. This process occurs in lymphocytes under several conditions, some of which appear to be genetically programmed (and hence are correctly cited as examples of 'programmed cell death').

and others are due to withdrawal of growth factors or as a consequence of antigen receptor-mediated activation [1]. During the development of T lymphocytes in the thymus, cells that express receptors that fail to recognize self MHC-encoded molecules, and are therefore not positively selected, undergo apoptotic programmed cell death. Immature T cells that express high-affinity receptors for self peptide + self MHC are believed to undergo activation-induced apoptosis as a result of antigen recognition. This is the principal mechanism of negative selection (clonal deletion) of self antigen-reactive thymocytes [2-4]. It can be mimicked by exposing immature T cells to antibodies that crosslink T cell receptors [5,6]. Similar mechanisms are believed to regulate the development of the mature, self-tolerant B-cell repertoire, although this process is not as well understood as it is for T lymphocytes [7].

After lymphocytes mature and leave the generative tissues (thymus and bone marrow), they are functionally competent, i.e. capable of responding to antigenic stimulation by proliferating and differentiating into effector cells. Among the progeny of antigen-stimulated lymphocytes, only a small fraction develop into functional effector cells and memory cells, and the majority probably die by apoptosis [8]. This is a process of activation-induced cell death that may be enhanced by the exposure of antigen-stimulated lymphocytes to growth factors, such as interleukin (IL)-2 in the case of T cells [9]. In mice, administration of large doses of anti-T cell receptor antibodies or superantigens, such as staphylococcal enterotoxins, results in the deletion of T cells that express antigen receptors which specifically bind these antibodies or superantigens [10-12]. This is probably also due to

Abbreviations

CTL—cytotoxic T lymphocyte; FasL—Fas ligand; *gld*—generalized lymphoproliferative disease; IL—interleukin;
lpr—lymphoproliferative gene; SLE—systemic lupus erythematosus; TNF—tumor necrosis factor.

Apoptosis, Fas and systemic autoimmunity: the MRL-*lpr/lpr* model

Gary G Singer, Ana C Carrera, Ann Marshak-Rothstein,
Carlos Martínez-A and Abul K Abbas

Brigham and Women's Hospital and Harvard Medical School, Boston, USA, Universidad
Autónoma de Madrid, Madrid, Spain and Boston University School of Medicine, Boston, USA

Proteins encoded by the *fas* and *fas ligand (fasl)* genes are involved in apoptotic cell death in lymphocytes. In this article we review the recent elucidation of the role of the Fas-FasL interactions in the maintenance of tolerance to self antigens and in the homeostatic regulation of lymphocyte clonal expansion, and discuss the mechanisms of autoimmunity in Fas- and FasL-deficient mutant mouse strains.

Current Opinion in Immunology 1994, 6:913-920

Introduction

The death of lymphocytes by apoptosis is an important mechanism for maintaining self-tolerance and homeostasis in the immune system. Failure of apoptotic cell death has long been postulated as a cause of loss of self-tolerance and the development of autoimmunity. Mice expressing the *lpr* (lymphoproliferation) and *gld* (generalized lymphoproliferative disease) mutations develop an accelerated, lupus-like systemic autoimmune disease. The findings that these single-gene defects are due to abnormalities in the cell death associated gene, *fas*, or in the Fas ligand (*FasL*), have formally established the importance of apoptotic cell death in preventing autoimmunity. These two animal models represent the first autoimmune diseases in which the major underlying genetic abnormality is known. In this paper, we review the role of apoptosis in the maintenance of self-tolerance, the genetic and immunologic basis of disease in MRL-*lpr/lpr* mice, and the relevance of this model in other animal and human models of systemic autoimmunity.

Lymphocyte apoptosis and tolerance to self antigens

Apoptotic cell death is characterized by fragmentation of DNA into nucleosome-sized fragments, nuclear dissolution and cell shrinkage. This process occurs in lymphocytes under several conditions, some of which appear to be genetically programmed (and hence are correctly cited as examples of 'programmed cell death').

and others are due to withdrawal of growth factors or as a consequence of antigen receptor-mediated activation [1]. During the development of T lymphocytes in the thymus, cells that express receptors that fail to recognize self MHC-encoded molecules, and are therefore not positively selected, undergo apoptotic programmed cell death. Immature T cells that express high-affinity receptors for self peptide + self MHC are believed to undergo activation-induced apoptosis as a result of antigen recognition. This is the principal mechanism of negative selection (clonal deletion) of self antigen-reactive thymocytes [2-4]. It can be mimicked by exposing immature T cells to antibodies that crosslink T cell receptors [5,6]. Similar mechanisms are believed to regulate the development of the mature, self-tolerant B-cell repertoire, although this process is not as well understood as it is for T lymphocytes [7].

After lymphocytes mature and leave the generative tissues (thymus and bone marrow), they are functionally competent, i.e. capable of responding to antigenic stimulation by proliferating and differentiating into effector cells. Among the progeny of antigen-stimulated lymphocytes, only a small fraction develop into functional effector cells and memory cells, and the majority probably die by apoptosis [8]. This is a process of activation-induced cell death that may be enhanced by the exposure of antigen-stimulated lymphocytes to growth factors, such as interleukin (IL)-2 in the case of T cells [9]. In mice, administration of large doses of anti-T cell receptor antibodies or superantigens, such as staphylococcal enterotoxins, results in the deletion of T cells that express antigen receptors which specifically bind these antibodies or superantigens [10-12]. This is probably also due to

Abbreviations

CTL—cytotoxic T lymphocyte; Fas—Fas ligand; *gld*—generalized lymphoproliferative disease; IL—interleukin; *lpr*—lymphoproliferative gene; SLE—systemic lupus erythematosus; TNF—tumor necrosis factor.

activation-induced apoptosis, and may be an exaggerated version of the phenomenon that normally occurs in clones of antigen-specific lymphocytes that encounter the antigen. Activation-induced cell death in mature lymphocytes is a homeostatic mechanism that functions to regulate the size of antigen-stimulated clones. It is not known whether the same mechanism is responsible for the maintenance of peripheral tolerance to tissue-specific self antigens, although recent findings in MRL-*lpr/lpr* mice indicate that apoptotic cell death may indeed be an important mechanism of peripheral self-tolerance.

Genetic control of apoptosis in lymphocytes

A large number of genes that determine apoptotic programmed cell death have been identified in *Caenorhabditis elegans* [13], and more recently in *Drosophila* sp. [14]. Much of the interest in these genes has focused on their role in embryogenesis and the development of specialized tissues, including the central nervous system. Although homologous genes are being identified in mammals, it appears that apoptosis in lymphocytes and other hemopoietic cells is regulated by unique genes. These include members of the tumor necrosis factor (TNF) receptor/*fas* superfamily, proto-oncogenes such as *bcl-2* and *c-myc*, and genes that regulate the cell cycle, such as *p53* [15].

The TNF receptor/*fas* superfamily

The *Fas* protein was first identified as a membrane protein and as a target for antibodies that induced apoptosis in cell lines [16]. The human protein called APO-1, which was also identified as a target for an antibody that induced apoptosis, is identical to *Fas* [17], as is CD95. *Fas* is a member of a family of proteins with 12 known members, including the p55 and p75 components of the TNF receptor, CD40, OX40, CD27 and CD30 [18⁺]. All members of the family are type I membrane proteins containing homologous extracellular domains with three to six cysteine-rich 40 amino acid long pseudorepeats, and cytoplasmic domains of variable length without significant sequence homology (Fig. 1). Most of these proteins can be produced in soluble forms, which are usually generated by proteolysis of the membrane proteins. The ligands for these proteins are all type II membrane molecules, with extracellular carboxy-terminal regions and intracellular amino termini (Fig. 1). Eight of these ligands have been cloned; sequence homologies among them are limited to the carboxyl terminal ~150 residues, which presumably form the receptor-binding regions. All the ligands can also be expressed in membrane-associated or secreted forms.

The *fas* gene encodes a ~45 kDa protein. It is expressed in activated mature T cells, thymus, liver, ovary, lung and heart [19]. *FasL* is a ~40 kDa protein; *FasL* RNA is present in testis, small intestines, kidney, lung, activated splenocytes, and, to a lesser degree, activated thy-

mocytes, but there is little information available about protein expression [20⁺,21].

Members of the TNF receptor/*fas* superfamily all regulate the growth, differentiation and apoptotic cell death of lymphocytes. Binding of *FasL* or anti-*Fas* antibodies to cell-surface *Fas* induces apoptosis, much like the binding of TNF to its receptor on target cells. The process of *Fas*-mediated cell death is essential for maintaining T-cell tolerance to self antigens, as revealed by analysis of the MRL-*lpr/lpr* mouse strain. In addition, apoptosis induced by cytolytic T lymphocytes (CTLs) is also mediated by *FasL* produced by the CTLs binding to *Fas* on target cells. This mechanism of target-cell killing complements osmotic lysis induced by the insertion of perforin into target cell membranes [22⁺]. In fact, the principal mechanism of cytotoxicity by CD4⁺ CTLs is *Fas*-mediated [23⁺]. In contrast to the *FasL*-*Fas* interaction, CD40L or anti-CD40 antibodies protect CD40-expressing B cells from death, and stimulate the growth and differentiation of these cells [24–26]. In some cells, TNF- α also induces proliferation and not death [27], and even *Fas* engagement has been shown to increase the proliferation of some T lymphocytes [28]. The mechanisms by which different TNF receptor/*fas* superfamily members transduce ligand-induced signals and the biochemical basis for the distinct functional consequences of ligation of these receptors are entirely unknown. The regulation of *Fas* and *FasL* expression in lymphocytes is also not yet fully defined, and this is an area of intensive investigation at present. Activated T helper (Th) 1 clones may express higher levels of *FasL* than most Th2 clones [29], but it is unlikely that *FasL* expression is restricted to any one T cell subset. How cytokines and costimulators influence *Fas* and *FasL* expression and function is another, largely unresolved issue.

Other genes that regulate apoptosis in lymphocytes

The proto-oncogene, *bcl-2*, promotes cell division largely by inhibiting programmed cell death. Transgenic mice that overexpress *bcl-2* in lymphoid cells show prolonged antibody responses to immunization, and enhanced generation of memory B cells [30,31]. Targeted disruption of the *bcl-2* gene results in a dramatic postnatal involution of both thymus and spleen, secondary to widespread apoptotic cell death [32]. *Fas*-mediated apoptosis may be counteracted by overexpression of *bcl-2* in cell lines, but the mechanism of this effect is not known [33]. The proto-oncogene *c-myc* generally inhibits apoptosis in cell lines. The tumor suppressor *p53* is required for apoptosis induced in thymocytes by ionizing radiation [34]. Recently, Nur 77, an orphan steroid receptor, has been shown to be required for activation-induced apoptosis in immature T cells [35,36]. Many of these apoptosis-regulating genes were first identified in tumors, and postulated to play critical roles in the prolonged survival and uncontrolled growth of neoplastic cells. Subsequent studies have implicated these same genes in the growth and survival of normal cells, but the importance of *bcl-*

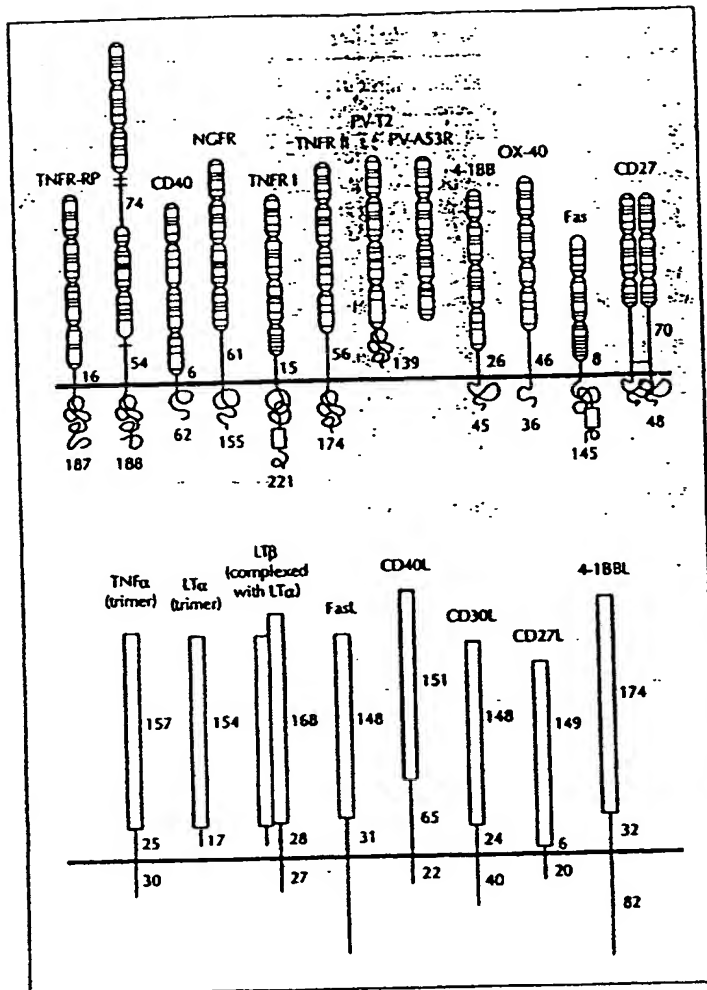


Fig. 1. Members of the TNF receptor/Fas superfamily (top) and the TNF/FasL superfamily (bottom). Published with permission [18**].

2, *c-myc*, *p53* and *Nur 77* in the physiologic regulation of immune responses and self-tolerance is not yet established.

Autoimmunity caused by the *lpr* and *gld* mutations

The spontaneous lupus-like autoimmune disease of MRL-*lpr/lpr* and *gld* mice has provided a powerful model for analyzing the mechanisms of self-tolerance and autoimmunity. Although considerable confusion has arisen about the basis of autoimmunity in these strains, recent molecular and immunologic analyses have provided important insights.

Immunologic abnormalities in MRL-*lpr/lpr* and *gld* disease MRL mice homozygous for the *lpr* gene (MRL-*lpr/lpr*) develop multiple autoantibodies, including antinuclear protein antibodies, rheumatoid factor, and immune complex nephritis, which is fatal by ~6 months [37]. In addition, after ~6 weeks of age, the mice develop progressive lymphadenopathy due to the accumulation of an unusual population of CD4⁺ CD8⁻ TCRαβ⁺ CD3⁺ (double-negative) T cells that also express the CD45R isoform called B220, normally a marker of the B-lymphocyte lineage. These double-negative T cells do not proliferate and are inert to extrinsic stimuli, including anti-CD3 antibodies and IL-2. One recent paper shows that the double-negative cells can be induced to proliferate when cultured with anti-CD3 and an activating antibody against CD28, TCR for co-stimulators [38], but

the significance of this observation is not known. Since the cells that cause lymphadenopathy are not actively proliferating, the term 'lymphoproliferation' is actually a misnomer. MRL-*lpr/lpr* mice also contain an abnormal number of autoreactive CD4⁺ T cells, which are detectable before the development of lymphadenopathy [39]. CD4⁺ T cell lines from MRL-*lpr/lpr* mice are resistant to cell death induced by high concentrations of anti-CD3 or anti-TCR antibodies [40,41,42], and these T cells have a striking growth advantage over normal CD4⁺ T cells in mixed cultures stimulated by alloantigens or other T-cell activators [43]. Autoreactive CD4⁺ T cells are capable of helping syngeneic B lymphocytes by inducing proliferation and antibody secretion, whereas the double-negative T cells are not functional in these assays (GG Singer, A Marshak-Rothstein, AK Abbas, unpublished data). It is not yet known whether particular cytokines play obligatory roles in the helper function of autoreactive CD4⁺ cells.

Mice homozygous for the *gld* gene develop a phenotypically very similar disease, characterized by autoantibody production, immune complex nephritis, lymphadenopathy due to the accumulation of double-negative T cells, and a high frequency of autoreactive CD4⁺ T lymphocytes [37,43]. The responses of *gld* T cells to extrinsic stimuli have not been analyzed in detail.

Several lines of evidence indicate that autoimmunity in MRL-*lpr/lpr* mice is dependent on CD4⁺ helper T cells, and can be segregated from the lymphadenopathy. Treatment of MRL-*lpr/lpr* mice with an antibody against CD4 prevents or retards disease [44]. In contrast, anti-CD8 antibody treatment prevents lymphadenopathy but not autoimmunity [45]. More definitive proof has come from breeding studies. MRL-*lpr/lpr* mice bred with mice deficient in class II MHC do not develop autoimmunity, probably because class II^{-/-} animals lack CD4⁺ T cells, but again lymphadenopathy is not affected [46]. The same result is seen in CD4^{-/-} MRL-*lpr/lpr* mice, whereas CD8^{-/-} MRL-*lpr/lpr* mice do develop autoantibodies and show variable degrees of lymphadenopathy (D Roh and TW Mak, personal communication). Thus, the conclusion of these analyses is that autoantibody production is dependent on autoreactive CD4⁺ helper T cells, whereas lymphadenopathy may be due to abnormalities in either CD4⁺ or CD8⁺ T cells (and is not, therefore, abolished in any of the mice described above). It is likely that the *lpr* mutation affects cell lineages other than CD4⁺ T lymphocytes. For instance, in chimeras between *lpr/lpr* and normal strains, B cells derived from the former have a striking growth advantage and are required for the development of autoimmunity, suggesting that *lpr/lpr* B cells are also intrinsically abnormal [47,48].

If CD4⁺ autoreactive T cells are critical for the development of autoimmunity, the key question that arises is why do MRL-*lpr/lpr* and *gld* mice contain large numbers of autoreactive Th lymphocytes?

Genetic basis of autoimmunity in MRL-*lpr/lpr* and *gld/gld* mice

The single most important advance in our understanding of autoimmunity in these strains has been the identification by Nagata's laboratory of the *lpr* mutation as a defect in the *fas* gene [49]. This abnormal *fas* gene contains a retrotransposon insertion in the second intron, leading to aberrant splicing and premature termination of transcription [50,51]. As a result, cells from MRL-*lpr/lpr* mice produce greatly reduced or undetectable *fas* transcripts. Although less is known about protein expression, the available data indicate that Fas protein is also essentially undetectable in *lpr/lpr* mice. Formal proof that the *fas* defect causes both autoimmunity and lymphadenopathy has come from the demonstration that the disease of MRL-*lpr/lpr* mice can be cured if a normal *fas* gene is expressed as a transgene in the T cells of these animals [52].

The cloning of the gene encoding the FasL [20,21] was rapidly followed by the finding that *gld* is a point mutation in the extracellular domain of the gene encoding FasL [53]. The identity of the *gld* gene has been confirmed by positional cloning [54]. Moreover, it has been shown that this abnormal FasL is incapable of inducing apoptosis in Fas-expressing target cells. The demonstration that *lpr* and *gld* are abnormalities in the genes encoding a receptor-ligand pair provides the structural explanation for the long-held view that these autoimmune strains are caused by abnormalities in a complementary ligand:receptor pair [55].

However, the disease of MRL-*lpr/lpr* mice, although clearly *fas*-mediated, is significantly modified by background genes. For instance, the levels of serum autoantibodies and the severity of nephritis vary depending on the background strain [37]. Furthermore, the MRL strain is itself autoimmune-prone, and the *lpr* defect markedly accelerates the development of autoimmunity.

Mechanisms of autoimmunity in MRL-*lpr/lpr* mice

On the basis of the known immunologic and genetic abnormalities in this strain, it is reasonable to postulate that a failure of apoptotic cell death in CD4⁺ T lymphocytes is responsible for the persistence of pathogenic autoreactive Th cells and the subsequent production of autoantibodies. Autoreactive T cells may persist either because of an abnormality in negative selection in the thymus or because of a peripheral defect. Recent data indicate that thymic selection, assessed by the expression of TCR Vβs, occurs normally in the thymus of MRL-*lpr/lpr* mice [56]. More direct evidence that the defect is peripheral and not thymic has come from *lpr/lpr* mice expressing a transgene-derived TCR specific for a known peptide + class II MHC. TCR-expressing CD4⁺ T cells develop normally in these mice but are resistant

to activation-induced apoptotic cell death. More importantly, the systemic administration of high doses of peptide results in comparable deletion of intrathymic T cells expressing the transgenic TCR in *lpr/lpr* and normal (+/+) mice, whereas mature T cells in peripheral lymphoid tissues are deleted in +/+ but not in *lpr/lpr* animals [57]. These results indicate that Fas plays an essential role in activation-induced death of mature T cells in the periphery, but not in negative selection in the thymus. Therefore, one can postulate that normally, some self antigen-reactive T cells may mature and enter peripheral tissues, where an encounter with self antigens leads to FasL/Fas-mediated apoptosis. In MRL-*lpr/lpr* mice, failure of apoptosis results in the persistence of these autoreactive T cells, which help autoreactive B cells that are not deleted. The B cells are thus stimulated to produce high-affinity autoantibodies (Fig. 2). The insensitivity of *lpr/lpr* B cells to Fas/Fas L-mediated cytotoxicity may also contribute to autoantibody production [58]. Promoting activation-induced cell death may restore peripheral tolerance even in Fas- or FasL-deficient mice. This is the likely mechanism by which infection of MRL-*lpr/lpr* mice containing IL-2, but not IL-4, or granulocyte-macrophage colony-stimulating factor, viral vectors cures autoimmunity [59,60], since IL-2 is known to program T cells for apoptosis [9]. Failure of peripheral activation-induced T cell death is also the probable explanation for the relative inefficiency of superantigens to delete specific T cells in MRL-*lpr/lpr* mice [61].

Although the model presented in Fig. 2 may be the simplest explanation for the development of autoimmunity in *lpr/lpr* (and, by inference, *gld/gld*) strains, many issues remain unresolved. First, the relationship between persistent autoreactive T cells and the accumulation of double-negative cells in lymphoid organs is not known. One possibility is that chronic stimulation of autoreactive T cells by self antigens leads to compensatory downmodulation of CD4 or CD8 co-receptors and accumulation of inert, 'double-negative' cells. Since this could happen in either CD4⁺ or CD8⁺ populations, selectively depleting either subset may not prevent lymphadenopathy. Alternatively, the double-negative T cells may arise from some abnormal thymic population that emigrates to the periphery [62]. The role of other cell surface molecules, such as CD2, in the accumulation of functionally anergic double-negative cells and in apoptosis in B cells has also been suggested [63,64], but its relevance to autoimmunity is unknown. Second, it is not clear why autoreactive T cells persist in an inappropriate way in *lpr/lpr* mice, but T-cell responses to immunization with foreign antigens in adjuvants do not show abnormally prolonged kinetics and are not aberrantly high (GG Singer, AK Abbas, unpublished data). Identifying the self antigens that elicit autoimmune reactions, and the way these antigens are presented to T cells, may provide important clues about the activation of autoreactive cells. Finally, the contributions of other components of the immune system, e.g. B cells, or of background genes, are not understood.

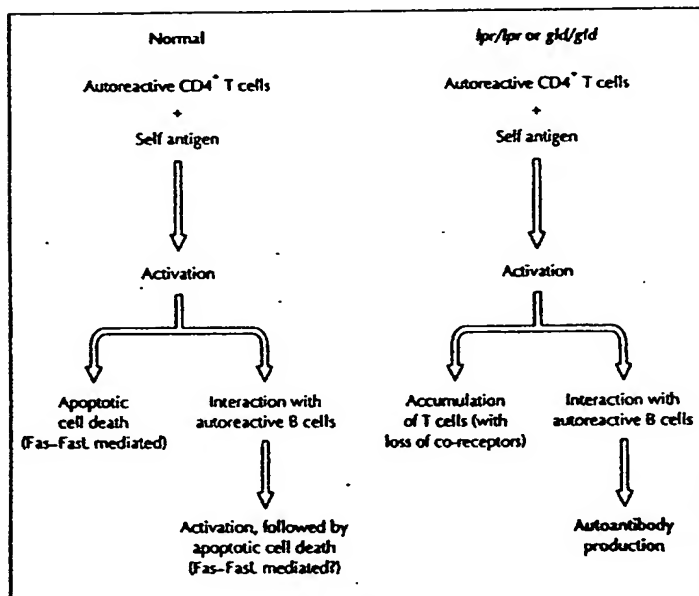


Fig. 2. A hypothetical model for the development of autoimmunity and lymphadenopathy as a result of the *lpr* and *gld* mutations. The postulated normal sequence of lymphocyte activation and regulation is shown for comparison.

Lymphocyte apoptosis and other models of autoimmunity

The relevance of the MRL-*lpr/lpr* model to human lupus and to other autoimmune disorders is an issue of great interest at present. Human systemic lupus erythematosus (SLE) is not caused by a single recessive gene, and is usually not associated with lymphadenopathy. By these criteria, SLE is not the same disease as the autoimmunity of MRL-*lpr/lpr* mice. Nevertheless, Mountz's group has recently found that some SLE patients have elevated serum levels of soluble Fas, which can block Fas-mediated apoptosis [65**]. This raises the possibility that secretion of soluble Fas may be the basis of autoimmunity in SLE, although the alternative, that high serum concentrations of soluble Fas are a result of lymphocyte activation, has not been excluded. Undoubtedly, many similar studies of SLE patients will be reported in the near future.

Regulating apoptotic cell death may be a general mechanism for inducing or ameliorating autoimmunity [15]. One line of transgenic mice constitutively expressing *bcl-2* in B lymphocytes develops an autoimmune syndrome very similar to that of MRL-*lpr/lpr* mice [30]. The likely mechanism is that overexpression of *bcl-2* prevents apoptotic cell death in self-reactive B lymphocytes. Conversely, repeated administration of a self antigen, myelin basic protein, reduces the autoimmune reactions specific to this protein, probably by activating T cells to produce large amounts of IL-2, and thus promoting apoptotic cell death [65**]. This raises the possibility that strategies for inducing apoptosis in lymphocytes may be of therapeutic benefit even in diseases associated with defects in the Fas-FasL pathway.

Conclusions

In summary, although MRL-*lpr/lpr* and *gld/gld* mice develop systemic disease due to multiple autoantibodies and immune complexes, they do not develop organ-specific, T cell mediated, autoimmune disorders. This suggests that different mechanisms may be responsible for maintaining tolerance to disseminated and tissue-specific self antigens. For instance, T-cell tolerance to widely disseminated self antigens may be due to negative selection in the thymus or Fas-dependent cell death in peripheral sites. Failure of the latter is the cause of the systemic disease of *lpr/lpr* and *gld/gld* mice. In contrast, tissue-restricted self antigens may normally be presented by co-stimulator deficient, antigen-presenting cells, and may thus induce anergy in potentially autoreactive T-cell clones that have attained maturity. This process of clonal anergy does not involve Fas-FasL interactions. Elucidation of the mechanisms of self-tolerance to diverse autoantigens will provide a rational basis for devising therapeutic strategies for spontaneous autoimmune diseases.

Acknowledgements

This work was supported by grants from the NIH (to AKA and AM-R) and CICyT, Comunidad Autonoma de Madrid (CAM), and Pharmacia (to CM-A). G Singer is a Fellow of the Medical Research Council of Canada.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Cohen B, Duke RC, Fadok VA, Sellins KS: Apoptosis and programmed cell death in immunity. *Ann Rev Immunol* 1992, 10:267-293.
 2. Murphy KM, Heimberger B, Loh DY: Induction by antigen of intrathymic apoptosis of CD4⁺CD8⁺TCR⁺ thymocytes. *In Vivo*. *Science* 1990, 250:1720-1723.
 3. Kappler JW, Roehm N, Marrack P: T cell tolerance by clonal elimination in the thymus. *Cell* 1987, 49:273-280.
 4. Kisielow P, Blüthmann H, Staerz UD, Steinmetz M, von Boehmer H: Tolerance in T-cell receptor transgenic mice involves deletion of nonmature CD4⁺8⁺ thymocytes. *Nature* 1988, 333:742-746.
 5. Smith CA, Williams G, Kingston R, Jenkinson EJ, Owen JF: Antibodies to CD3/T-cell receptor complex induce death by apoptosis in immature T cells in thymic cultures. *Nature* 1989, 337:181-184.
 6. Shi YF, Bissonnette RP, Parfrey N, Szalay M, Kubo RT, Green DR: *In vivo* Administration of monoclonal antibodies to the CD3 T cell receptor complex induces cell death (apoptosis) in immature thymocytes. *J Immunol* 1991, 146:3340-3346.
 7. Goodnow CC: Transgenic mice and analysis of B-cell tolerance. *Annu Rev Immunol* 1992, 10:489-518.
 8. Sprent J: T and B memory cells. *Cell* 1994, 76:315-322.
 9. Lenardo MJ: Interleukin-2 programs mouse αβ T lymphocytes for apoptosis. *Nature* 1991, 353:858-861.
 10. Kawabe Y, Ochi A: Programmed cell death and extrathymic reduction of VB8⁺ CD4⁺ T cells in mice tolerant to *Staphylococcus aureus* enterotoxin B. *Nature* 1991, 349:245-248.
 11. McCormack JE, Callahan JE, Kappler J, Marrack P: Profound deletion of mature T cells *in vivo* by chronic exposure to exogenous superantigen. *J Immunol* 1993, 150:3785-3792.
 12. Gonzalo JA, Baixeras E, Gonzalez-Garcia A, Goerge-Chandy A, van Rooijen N, Martinez-A C, Kroemer G: Differential *in vivo* effects of a superantigen and an antibody targeted to the same T cell receptor: activation induced cell death versus passive macrophage-dependent deletion. *J Immunol* 1994, 152:1597-1608.
 13. Driscoll M: Molecular genetics of cell death in the nematode *Caenorhabditis elegans*. *J Neurobiol* 1992, 23:1327-1351.
 14. White K, Grether ME, Abrams JM, Young L, Farrell K, Steller H: Genetic control of programmed cell death in *Drosophila*. *Science* 1994, 264:677-683.
 15. Kroemer G, Martinez-A C: Pharmacological inhibition of apoptosis. *Immunol Today* 1994, 23:235-242.
 16. Itoh N, Yonehara S, Ishii A, Yonehara M, Mizushima S-I, Sameshima M, Hase A, Seto Y, Nagata S: The polypeptide encoded by the cDNA for human cell surface antigen Fas can mediate apoptosis. *Cell* 1991, 66:233-243.
 17. Oehm A, Behrmann I, Falk W, Pawlitz M, Maier G, Klas C, Li-Weber M, Richards S, Oheim L, Trauth BC, et al: Purifica-

- tion and molecular cloning of the APO-1 cell surface antigen, a member of the tumor necrosis factor/nerve growth factor receptor superfamily. *J Biol Chem* 1992, 267:10709-10715.
18. Smith C A, Farrar T, Goodwin R G: The TNF receptor superfamily of cellular and viral proteins: activation, costimulation, and death. *Cell* 1994, 76:959-962.
- This is a thoughtful brief review of the proteins of the TNF receptor/Fas superfamily and their ligands. Conservation of structure and function is emphasized.
19. Watanabe-Fukunaga R, Brannan C I, Itoh N, Yonehara S, Copeland NG, Jenkins NA, Nagata S: The cDNA structure, expression, and chromosomal assignment of the mouse Fas antigen. *J Immunol* 1992, 148:1274-1279.
20. Suda T, Takahashi T, Golstein P, Nagata S: Molecular cloning and expression of the Fas ligand, a novel member of the tumor necrosis factor family. *Cell* 1993, 75:1169-1178.
- The search for the Fas ligand achieved the proportions of a 'holy grail', and culminated in this elegant description of the expression cloning of the ligand. Predictably, the Fas ligand is homologous to TNF, and induces apoptosis in Fas-expressing target cells.
21. Suda T, Nagata S: Purification and characterization of the Fas-ligand that induces apoptosis. *J Exp Med* 1994, 179:873-879.
22. Kojima H, Shinohara N, Hanaoka S, Someya-Shiota Y, Takagaki Y, Ohno H, Saito T, Katayama T, Yagita H, Okumura K, et al.: Two distinct pathways of specific killing revealed by perforin mutant cytotoxic T lymphocytes. *Immunity* 1994, in press.
- Fas/Fas-mediated apoptosis and perforin-mediated osmotic lysis are the principal mechanisms of killing by CTL, as revealed in this study of CTL from perforin-deficient mice.
23. Ju S-T, Cui HL, Panka DJ, Ettinger R, Marshak-Rothstein A: Participation of target Fas protein in apoptosis pathway induced by CD4⁺ Th1 and CD8⁺ cytotoxic T cells. *Proc Natl Acad Sci USA* 1994, 91:4185-4189.
- This study reports that the Fas protein is the principal target of CD4⁺ CTL, the cytolytic activity of which is inhibited by a soluble Fas-immunoglobulin fusion protein and is not seen with lpr/lpr target cells.
24. Liu Y-L, Johnson GD, Gordon I, MacLennan ICM: Germinal centres in T-cell-dependent antibody responses. *Immunol Today* 1992, 13:17-21.
25. Foy TM, Shepherd DM, Durie FH, Aruffo A, Ledbetter JA, Noelle RJ: In vivo CD40-gp39 interactions are essential for thymus-dependent humoral immunity. II. Prolonged suppression of the humoral immune response by an antibody to the ligand for CD40, gp39. *J Exp Med* 1993, 178:1567-1575.
26. Tsubata T, Wu L, Honjo T: B-cell apoptosis induced by antigen receptor crosslinking is blocked by a T-cell signal through CD40. *Nature* 1993, 364:645-648.
27. Hernández-Caselles T, Stutman O: Immune functions of tumor necrosis factor. I. Tumor necrosis factor induces apoptosis of mouse thymocytes and can also stimulate or inhibit IL-6-induced proliferation depending on the concentration of mitogenic costimulation. *J Immunol* 1993, 151:3999-4012.
28. Alderson MR, Arnáiz RJ, Marshakovsky E, Tough TW, Roux E, Schooley K, Ramsdell F, Lynch DH: Fas transduces activation signals in normal human T lymphocytes. *J Exp Med* 1993, 178:2231-2235.
29. Ramsdell F, Seaman MS, Miller RE, Picha KS, Kennedy MK, Lynch DH: Differential ability of Th1 and Th2 T cells to express Fas ligand and to undergo activation-induced cell death. *Int Immunol* 1994, in press.
30. Strasser A, Wittingham S, Vaux DL, Bath ML, Adams JM, Cory S, Harris AW: Enforced bcl-2 expression in B-lymphoid cells prolongs antibody responses and elicits autoimmune disease. *Proc Natl Acad Sci USA* 1991, 88:8661-8665.
31. Senman CL, Shutter JR, Hockenbery O, Kanagawa O, Korsmeyer SJ: bcl-2 inhibits multiple forms of apoptosis but not negative selection in thymocytes. *Cell* 1991, 67:879-888.
32. Veis DI, Sorenson CM, Shutter JR, Korsmeyer SJ: Bcl-2-deficient mice demonstrate fulminant lymphoid apoptosis, polycystic kidneys, and hypopigmented hair. *Cell* 1993, 75:229-240.
33. Itoh N, Tsujimoto Y, Nagata S: Effect of bcl-2 on Fas antigen-mediated cell death. *J Immunol* 1993, 151:621-627.
34. Lowe SW, Schmitt EM, Smith SW, Osborne BA, Jacks T: p53 is required for radiation-induced apoptosis in mouse thymocytes. *Nature* 1993, 362:847-852.
35. Liu ZG, Smith S, McLaughlin KA, Schwartz LM, Osborne BA: Apoptotic signals delivered through T cell receptors require the immediate early gene Nur 77. *Nature* 1994, 367:2821-2824.
36. Woronicz JD, Calnan B, Ngo V, Winoto A: Requirement for the orphan steroid receptor Nur 77 in apoptosis of T-cell hybridomas. *Nature* 1994, 367:277-281.
37. Cohen PL, Eisenberg RA: lpr and gld: Single gene models of systemic autoimmunity and lymphoproliferative disease. *Annu Rev Immunol* 1991, 9:243-269.
38. Giese T, Allison JP, Davidson WF: Functionally anergic lpr and gld B220⁺ T cell receptor (TCR)-α/β⁺ double-negative T cells express CD28 and respond to costimulation with phorbol myristate acetate and antibodies to CD28 and the TCR. *J Immunol* 1993, 151:597-609.
39. Weston KM, Ju S-T, Liu CY, Sy M-S: Autoreactive T cells in MRL/lpr/lpr mice. Characterization of the lymphokines produced and analysis of antigen-presenting cells required. *J Immunol* 1988, 141:1941-1946.
40. Russell JH, Rush B, Weaver C, Wang R: Mature T cells of autoimmune lpr/lpr mice have a defect in antigen-stimulated suicide. *Proc Natl Acad Sci USA* 1993, 90:4409-4413.
- This study reports that mature T cells from lpr/lpr mice are resistant to cell death induced by crosslinking of the antigen receptor.
41. Bossu P, Singer GG, Andres P, Ettinger R, Marshak-Rothstein A, Abbas AK: Mature CD4⁺ T lymphocytes from MRL/lpr mice are resistant to receptor-mediated tolerance and apoptosis. *J Immunol* 1993, 151:7233-7239.
- This study reports that CD4⁺ T cell clones from lpr/lpr mice are resistant to apoptosis and functional anergy induced by exposure to high concentrations of anti-CD3 antibody.
42. Gillette-Ferguson I, Sidman CL: A specific intercellular pathway of apoptotic cell death is defective in the mature peripheral T cells of autoimmune lpr and gld mice. *Eur J Immunol* 1994, 24:1181-1185.
- This study reports that mature T cells from lpr/lpr and gld/gld mice are not killed by culture with anti-CD3 antibody.
43. Ettinger R, Wang JK M, Bossu P, Papa K, Sidman CL, Abbas AK, Marshak-Rothstein A: Functional distinctions between MRL-lpr and MRL-gld lymphocytes. Normal cells reverse the gld but not lpr immunoregulatory defect. *J Immunol* 1994, 152:1557-1568.
44. Jabs DA, Kuppers RC, Saboori AM, Burek CL, Enger C, Lee B, Prendergast RA: Effects of early and late treatment with anti-CD4 monoclonal antibody on autoimmune disease in MRL/lpr/lpr mice. *Cell Immunol* 1994, 154:66-76.
45. Giese T, Davidson WF: Chronic treatment of C3H-lpr/lpr and C3H-gld/gld mice with anti-CD8 monoclonal antibody prevents the accumulation of double negative T cells but not autoantibody production. *J Immunol* 1994, 152:2000-2010.
46. Jevnikar AM, Grusby MJ, Glimcher LH: Prevention of nephritis in MHC class II-deficient MRL-lpr mice. *J Exp Med* 1994, 179:1137-1143.
- This study provides evidence that autoimmune disease can be dissociated from lymphadenopathy in MHC class II-deficient MRL-lpr mice.
47. Sobel ES, Katagiri T, Katagiri K, Morris SC, Cohen PL, Eisenberg RA: An intrinsic B cell defect is required for the production of autoantibodies in the lpr model of murine systemic autoimmunity. *J Exp Med* 1991, 173:1441-1449.
48. Nemazee D, Guet C, Buerki K, Marshak-Rothstein A: B lymphocytes from autoimmune-prone mouse strain MRL/lpr manifest an intrinsic defect in tetraparental MRL/lpr x DBA/2 chimeras. *J Immunol* 1991, 147:2536-2539.
49. Watanabe-Fukunaga R, Brannan CI, Copeland NG, Jenkins NA, Nagata S: Lymphoproliferative disorder in mice explained by

- defects in Fas antigen that mediates apoptosis. *Nature* 1992, 356:314-317.
50. Adachi M, Watanabe-Fukunaga R, Nagata S: Aberrant transcription caused by the insertion of an early transposable element in an intron of the Fas antigen gene of *lpr* mice. *Proc Natl Acad Sci USA* 1993, 90:1756-1760.

The demonstration that the *lpr* mutation is a retrotransposon insertion that leads to premature termination of transcription of the *Fas* gene.

 51. Chu J-L, Drappa J, Parnassa A, Elkon KB: The defect in Fas mRNA expression in MRL/lpr mice is associated with insertion of the retrotransposon. *ETN. J Exp Med* 1993, 178:723-730.

This study reports essentially the same finding as the previous reference and was published almost simultaneously.

 52. Wu J, Zhou T, Zhang J, He J, Cause WC, Mountz JD: Correction of accelerated autoimmune disease by early replacement of the mutated *lpr* gene with the normal Fas apoptosis gene in the T cells of transgenic MRL-lpr/lpr mice. *Proc Natl Acad Sci USA* 1994, 91:2344-2348.

One of Koch's postulates was fulfilled by this demonstration that expression of a normal *Fas* gene in *lpr/lpr* mice cures the autoimmune disease.

 53. Takahashi T, Tanaka M, Brannan CI, Jenkins NA, Copeland NG, Suda T, Nagata S: Generalized lymphoproliferative disease in mice, caused by a point mutation in the Fas ligand. *Cell* 1994, 76:969-976.

Yet another tour-de-force from Nagata's laboratory who identified the *gld* mutation as a point mutation in the *Fas* ligand gene, which results in an inability of the mutant FasL to induce apoptosis in Fas-expressing cell lines.

 54. Lynch DH, Watson ML, Alderson MR, Baum PR, Miller RE, Tough T, Gibson M, Davis-Smith T, Smith CA, Hunter K, et al.: The mouse Fas-ligand gene is mutated in *gld* mice and is part of a TNF family gene cluster. *Immunity* 1994, 1:131-136.

This study provides confirmation that *gld* is a point mutation in the *Fas* gene. By using a positional cloning approach, the authors reached the same conclusions as in [53**].

 55. Allen RD, Marshall JD, Roths JB, Sidman CL: Differences defined by bone marrow transplantation suggest that *lpr* and *gld* are mutations of genes encoding an interacting pair of molecules. *J Exp Med* 1990, 172:1367-1375.
 56. Herron LR, Eisenberg RA, Roper E, Kakkanaiah VN, Cohen PL, Kotzin BL: Selection of the T cell receptor repertoire in *lpr* mice. *J Immunol* 1993, 151:3450-3459.

Expression of T cell receptor V β genes is normal in thymocytes of *lpr/lpr* mice, indicating that superantigen-induced negative selection of thymocytes is not influenced by the deficiency of Fas.

 57. Singer CG, Abbas AK: The Fas antigen is involved in peripheral but not thymic deletion of T lymphocytes in T cell receptor transgenic mice. *Immunity* 1994, 1: in press.

In *lpr/lpr* mice expressing a TCR of known peptide specificity, administration of the peptide causes deletion of thymocytes but not of mature T cells. This paper provides formal support for the hypothesis that the Fas-FasL interaction is involved in activation-induced death of mature T cells but not in clonal deletion of developing thymocytes.

 58. Vignaux F, Colstien P: Fas-based lymphocyte-mediated cytotoxicity against syngeneic activated lymphocytes: a regulatory pathway? *Eur J Immunol* 1994 24:923-927.
 59. Kroemer G, Andreu JL, Gonzalo JA, Gutierrez-Ramos JC, Martinez-A C: Interleukin-2, autotolerance, and autoimmunity. *Adv Immunol* 1991, 50:147-235.
 60. Martinez-A C: The role of cytokines in the pathogenesis of autoimmunity. *Eur J Clin Invest* 1994, in press.
 61. Scott DE, Kirsch WJ, Steinberg AD: Studies of T cell deletion and T cell anergy following in vivo administration of SEB to normal and lupus-prone mice. *J Immunol* 1993, 150:664-672.
 62. Martinez-A C, Marcos MA, de Alborán IM, Alonso JM, de Cid R, Kroemer G, Coutinho A: Functional double-negative T cells in the periphery express TCR V β gene products that are deleted in single-positive T cells. *Eur J Immunol* 1993, 23:250-253.
 63. Clements JL, Wolfe J, Cooper SM, Budd RC: Reversal of hyporesponsiveness in *lpr* CD4-CD8- T cells is achieved by induction of cell cycling and normalization of CD2 and p59 lym expression. *Eur J Immunol* 1994, 24:558-565.
 64. Genaro AM, Gonzalo JA, Bosca L, Martinez-A C: CD2-CD48 interactions prevent apoptosis in murine B lymphocytes by upregulating Bcl-2 expression. *Eur J Immunol* 1994, in press.
 65. Cheng J, Zhou T, Liu C, Shapiro RP, Brauer ML, Kieffer MC, Barr PI, Mountz JD: Protection from Fas-mediated apoptosis by a soluble form of the Fas molecule. *Science* 1994, 263:1759-1762.

A soluble form of the Fas protein acts as a competitive inhibitor of Fas-mediated apoptosis. More interestingly, some patients with lupus exhibit high serum levels of soluble Fas; it remains to be determined whether this putative antagonist causes autoimmunity or is produced as a consequence of unregulated lymphocyte activation.

 66. Critchfield JM, Racke MK, Zühlig-Pflücker JC, Cannella B, Raine CS, Goverman J, Lenardo MJ: T cell deletion in high antigen dose therapy of autoimmune encephalomyelitis. *Science* 1994, 263:1139-1143.

GG Singer, Immunology Research Division, Department of Pathology, and Renal Division, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA 02115, USA.

AK Abbas, Immunology Research Division, Department of Pathology, Brigham and Women's Hospital and Harvard Medical School, Boston, MA 02155, USA.

AC Carrera and C Martínez-A, Centro Nacional de Biotecnología, Universidad Autónoma de Madrid, Campus de Cantoblanco, 28049 Madrid, Spain.

A Marshak-Rothstein, Department of Microbiology and Immunology, Boston University, School of Medicine, Boston, MA 02109, USA.

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ BLACK BORDERS
- ☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES
- ☒ FADED TEXT OR DRAWING
- ☒ BLURRED OR ILLEGIBLE TEXT OR DRAWING
- ☐ SKEWED/SLANTED IMAGES
- ☐ COLOR OR BLACK AND WHITE PHOTOGRAPHS
- ☐ GRAY SCALE DOCUMENTS
- ☒ LINES OR MARKS ON ORIGINAL DOCUMENT
- ☒ REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY
- ☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.